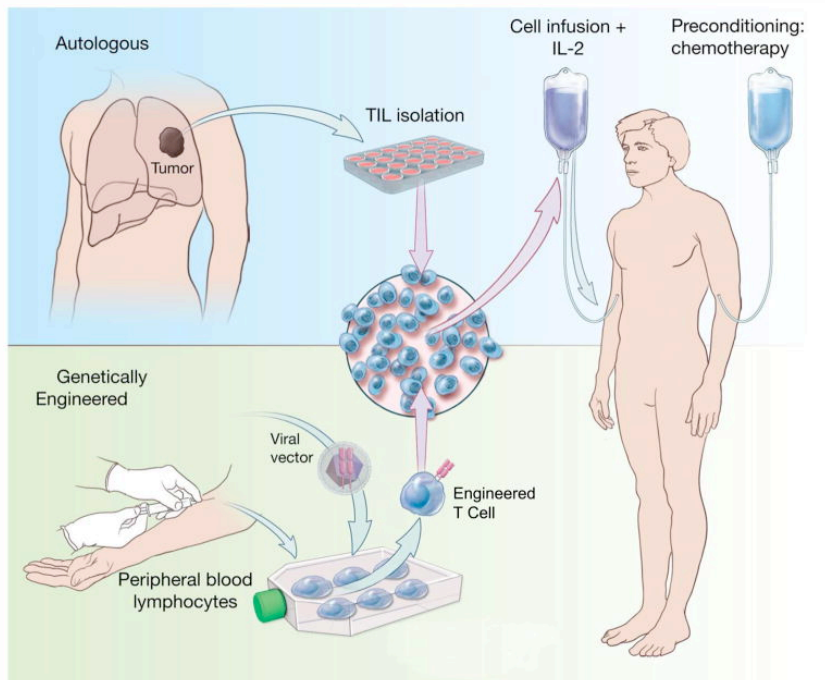


TCR gene therapy: clinical experience, toxicity and future perspectives

John Haanen



NETHERLANDS
CANCER
INSTITUTE

ANTONI VAN LEEUWENHOEK



Adoptive T-cell therapy

- Adoptive T-cell therapy for melanoma has proven efficacy
 - **TIL** (Dudley et al. Science 2002; Besser et al., Clin Cancer Res 2013; Joseph et al., Clin Cancer Res 2011; Pilon-Thomas et al., J Immunotherapy 2012; Donia et al., J Invest Dermatol 2013)
 - **Peripheral blood derived T-cells** (Mackensen et al., J Clin Oncol 2006; Yee et al., PNAS 2002; Hunder et al., NEJM 2008; Wallen et al., Plos One 2009; Khammari et al. J Invest Dermat 2009; Verdegaal et al., Cancer Immunol Immunother 2011)

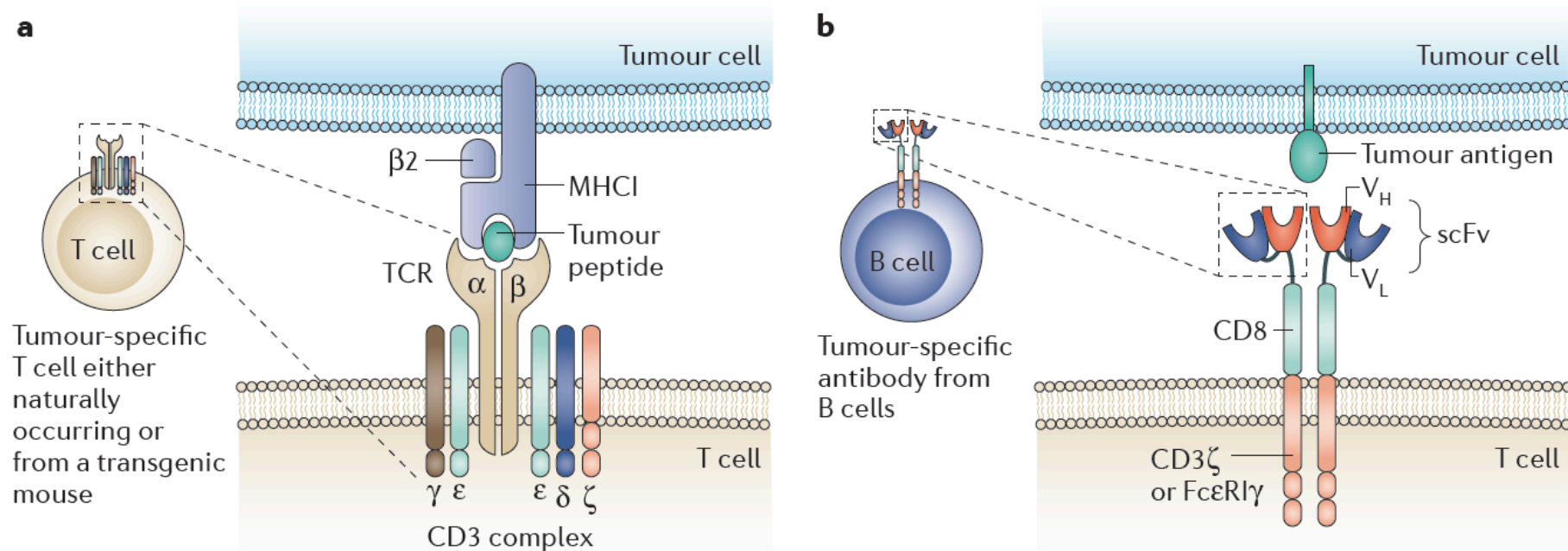
Why TCR gene therapy?

Alternative strategies for adoptive cell transfer as TIL therapy has limitations

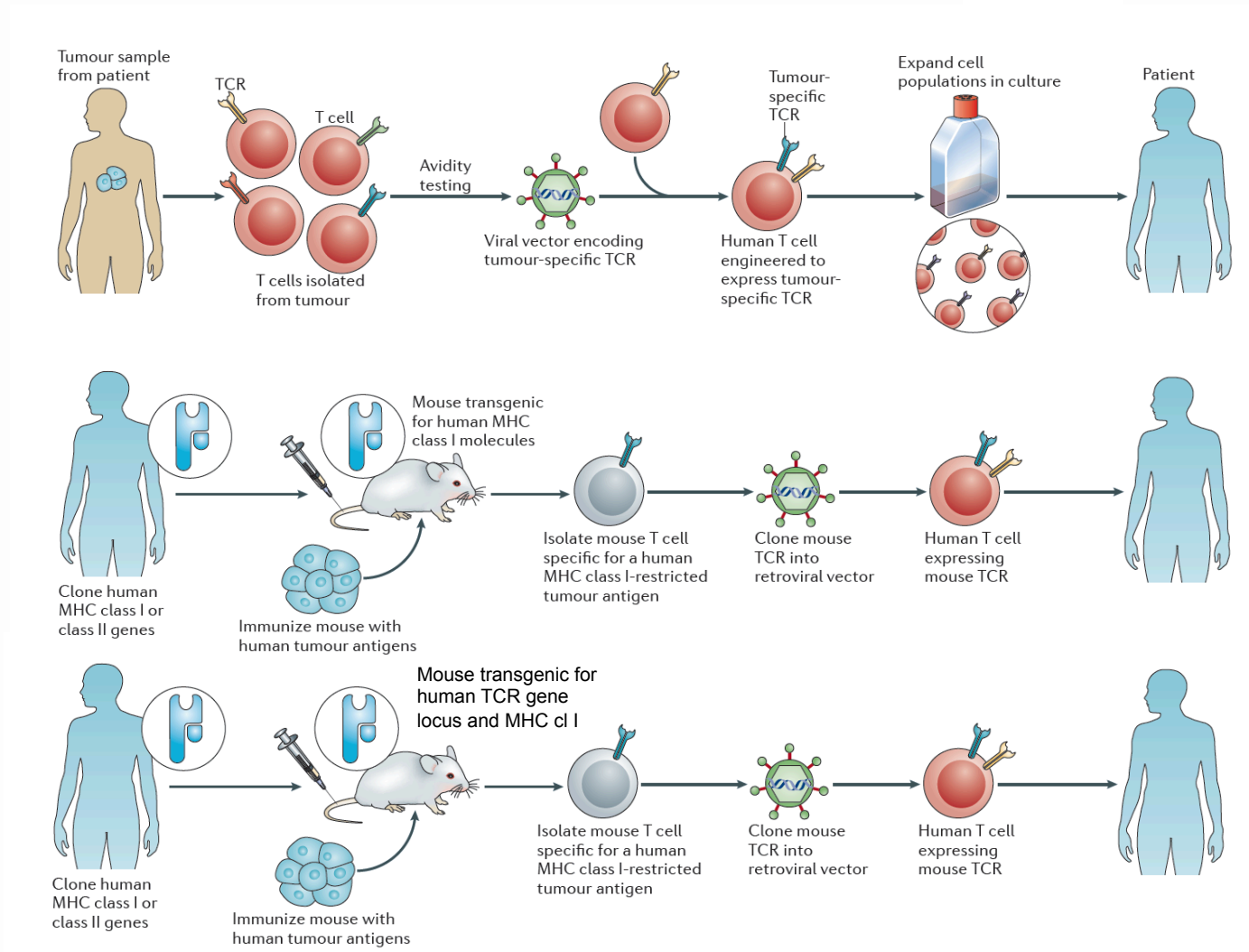
- Presence of resectable metastasis is required
- TIL grow from ~80-90% of resected tumor samples
- Very patient-specific treatment
- Laborious and time-consuming
- Very difficult to obtain tumor-specific TIL from tumors other than melanoma

Aim to generate off-the-shelf reagents for many cancer patients

Gene-modified T cells



Genetically modified peripheral blood lymphocytes



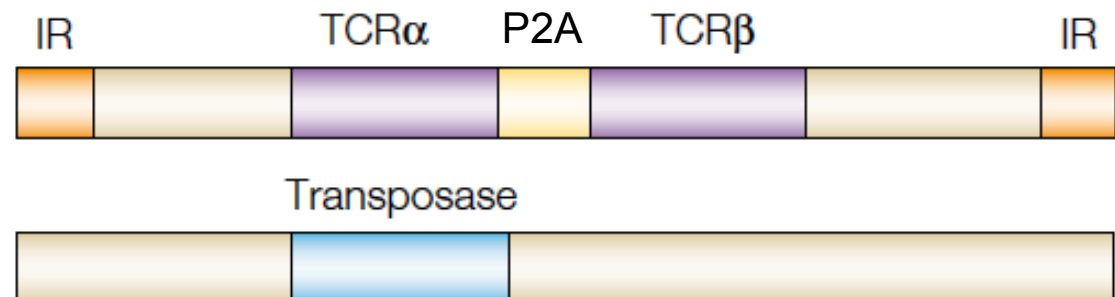
Modified from: Restifo et al., Nature Rev Immunol (2012)

Transduction platforms for gene transfer

a Retro- or lentiviral vector



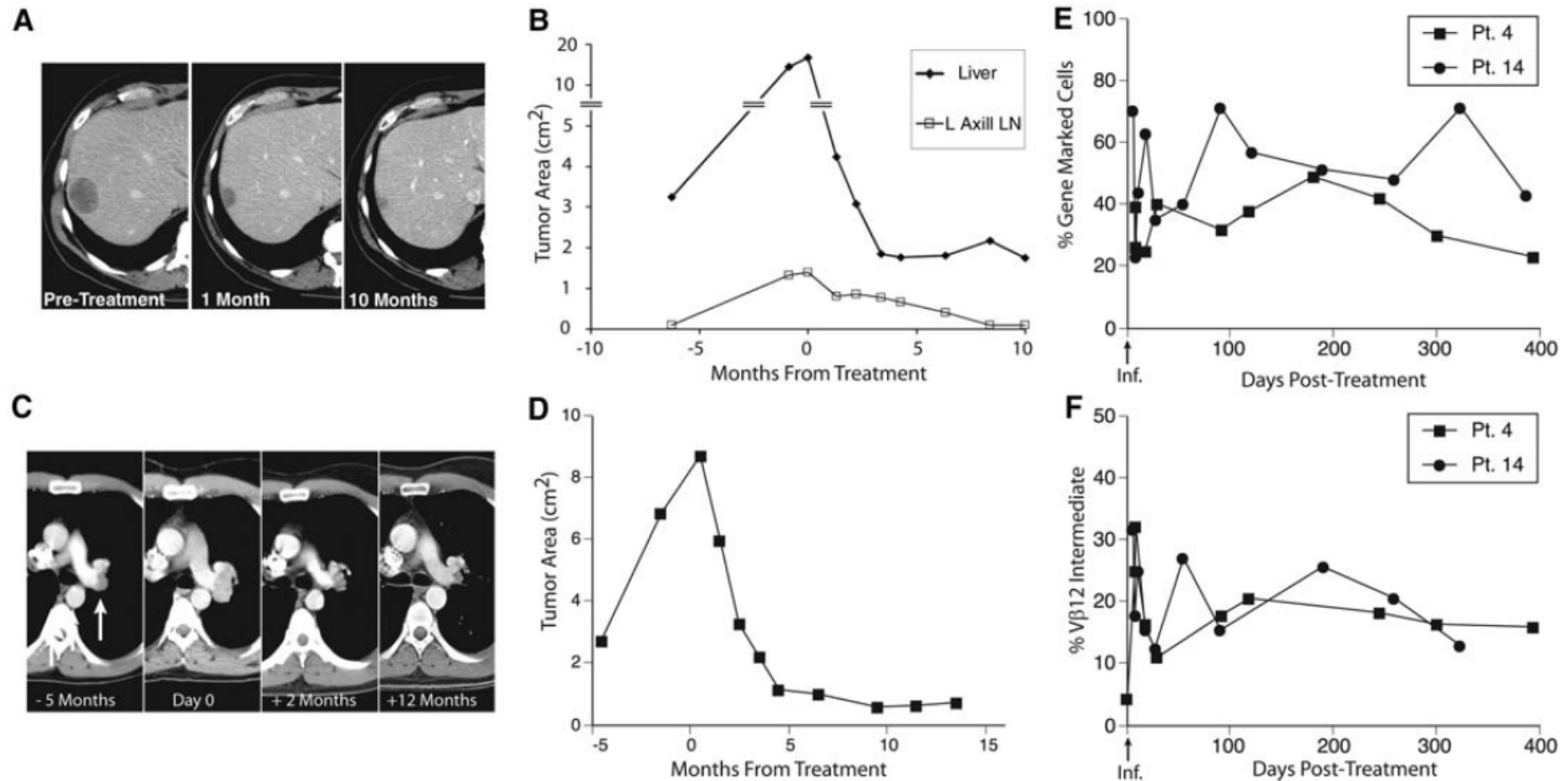
b Transposon-based vector



Clinical experience with TCR gene therapy

- 2006: MART-1 TCR gene therapy for melanoma
 - RR 13% (n=15)
 - (Morgan et al., Science 2006)

Cancer regression upon transfer of MART-1 TCR redirected T cells

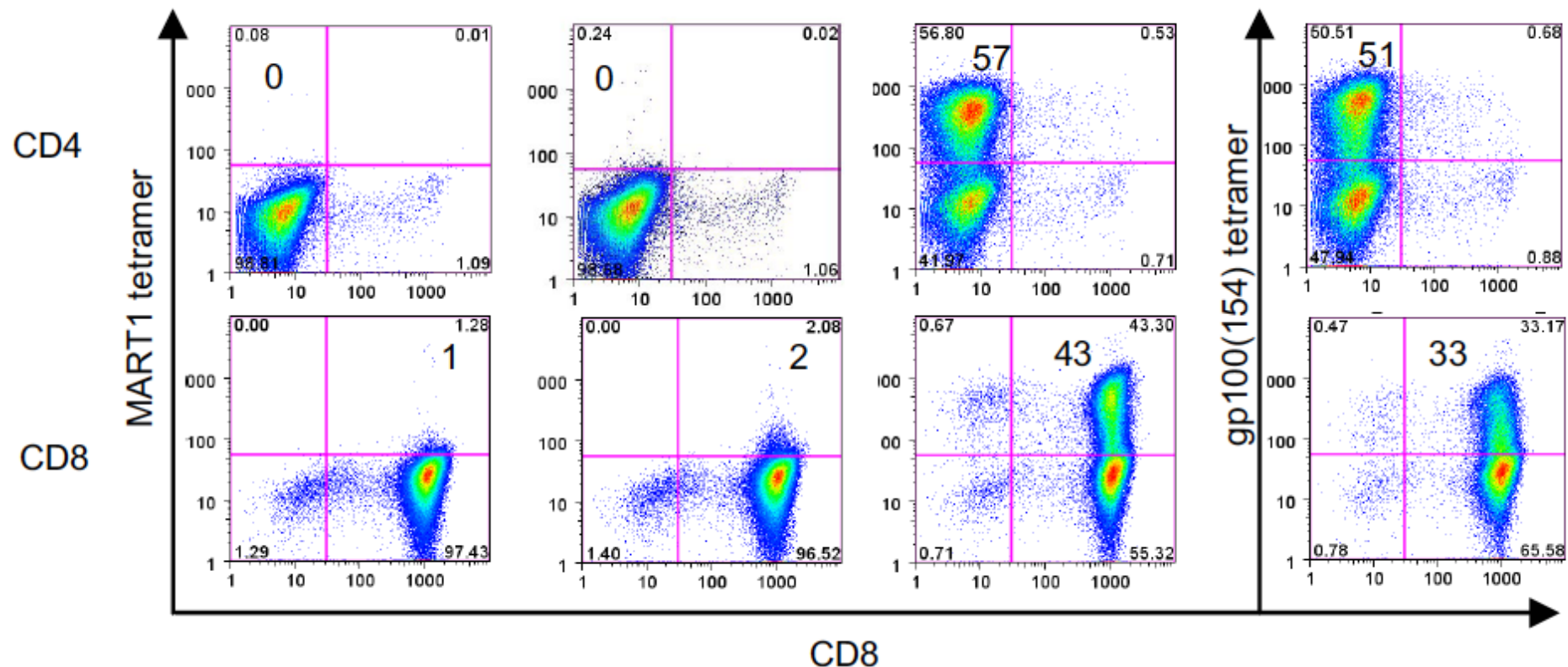


Morgan et al., Science 2006

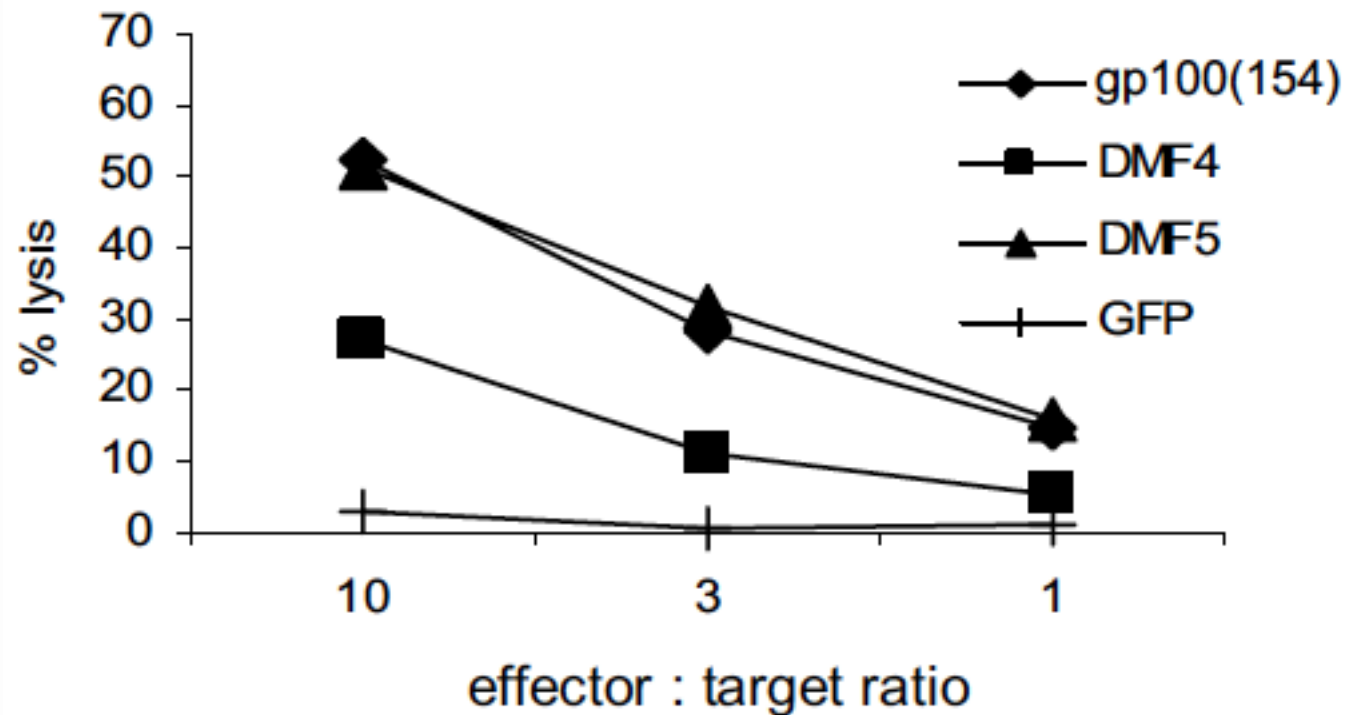
Clinical experience with TCR gene therapy

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- 2009: MART-1 and gp100 TCR gene therapy
 - RR 30% (MART-1 TCR; n=20)
 - RR 19% (murine gp100 TCR; n=16)
(Johnson et al., Blood 2009)

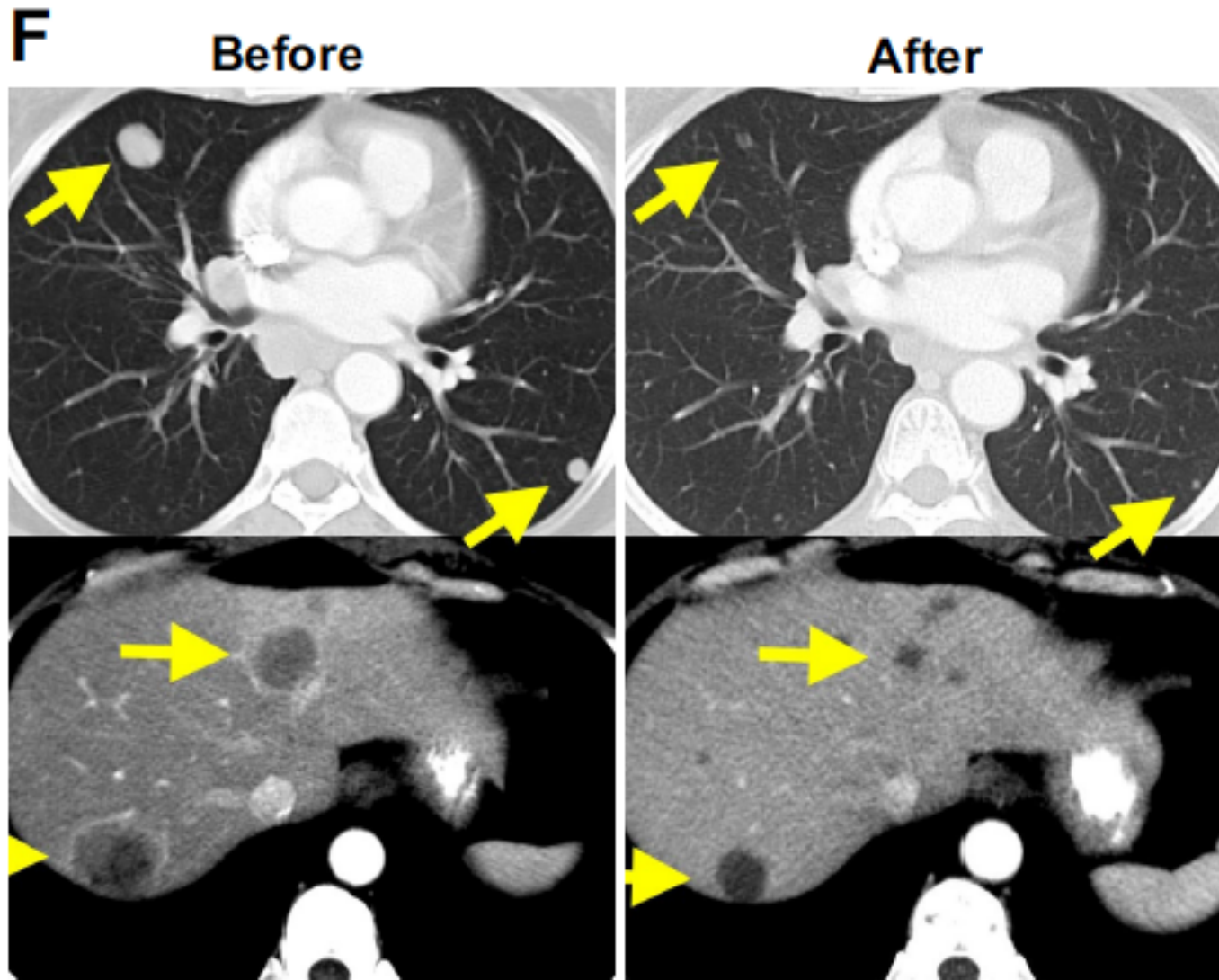
DMF5 and gp100 specific TCR were highly expressed by transduced CD4 and CD8 T cells



Higher cytolytic activity of 2nd generation MDA-specific TCRs



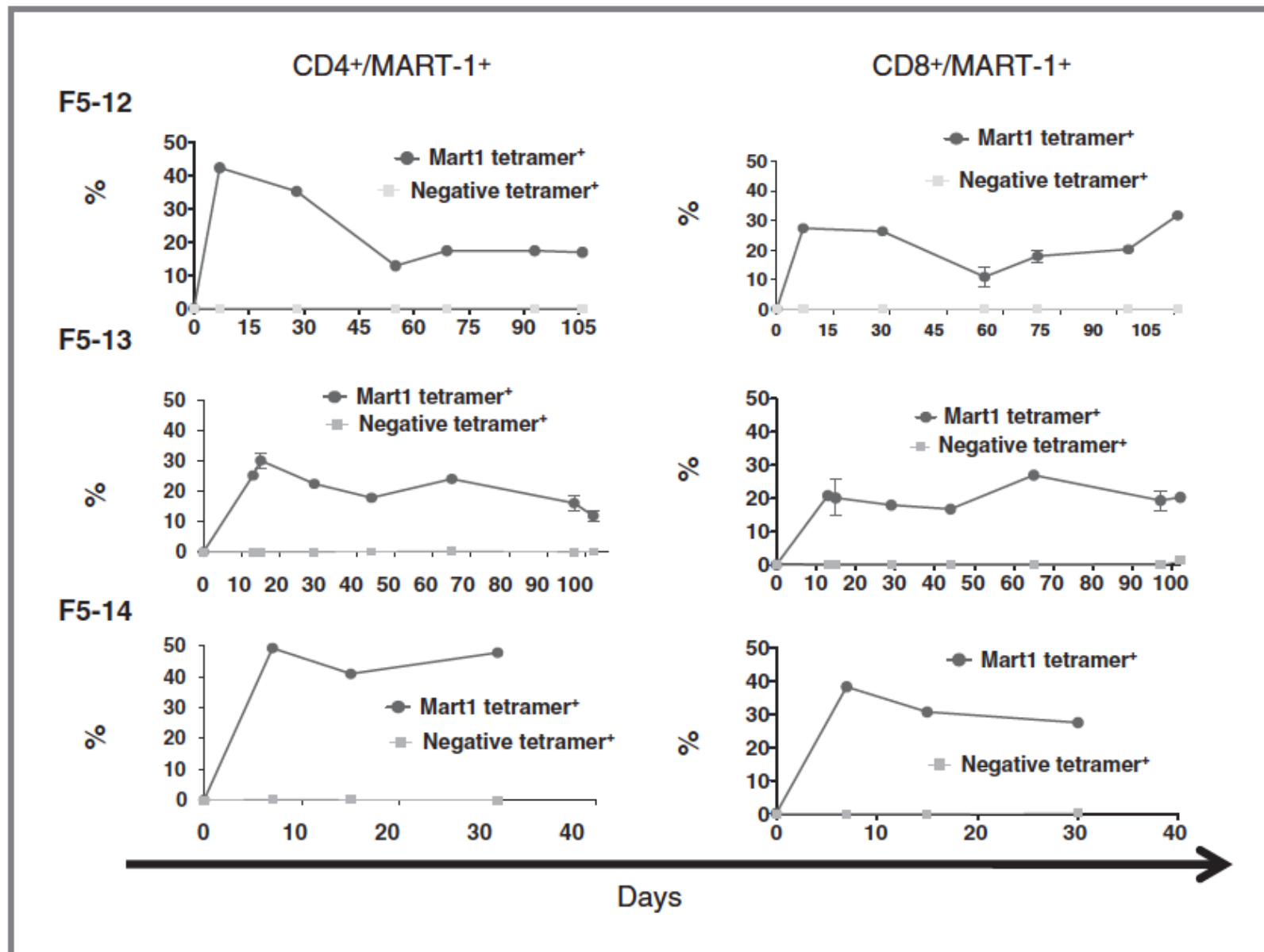
Clinical activity of MART-1 and gp100-specific TCR gene therapy



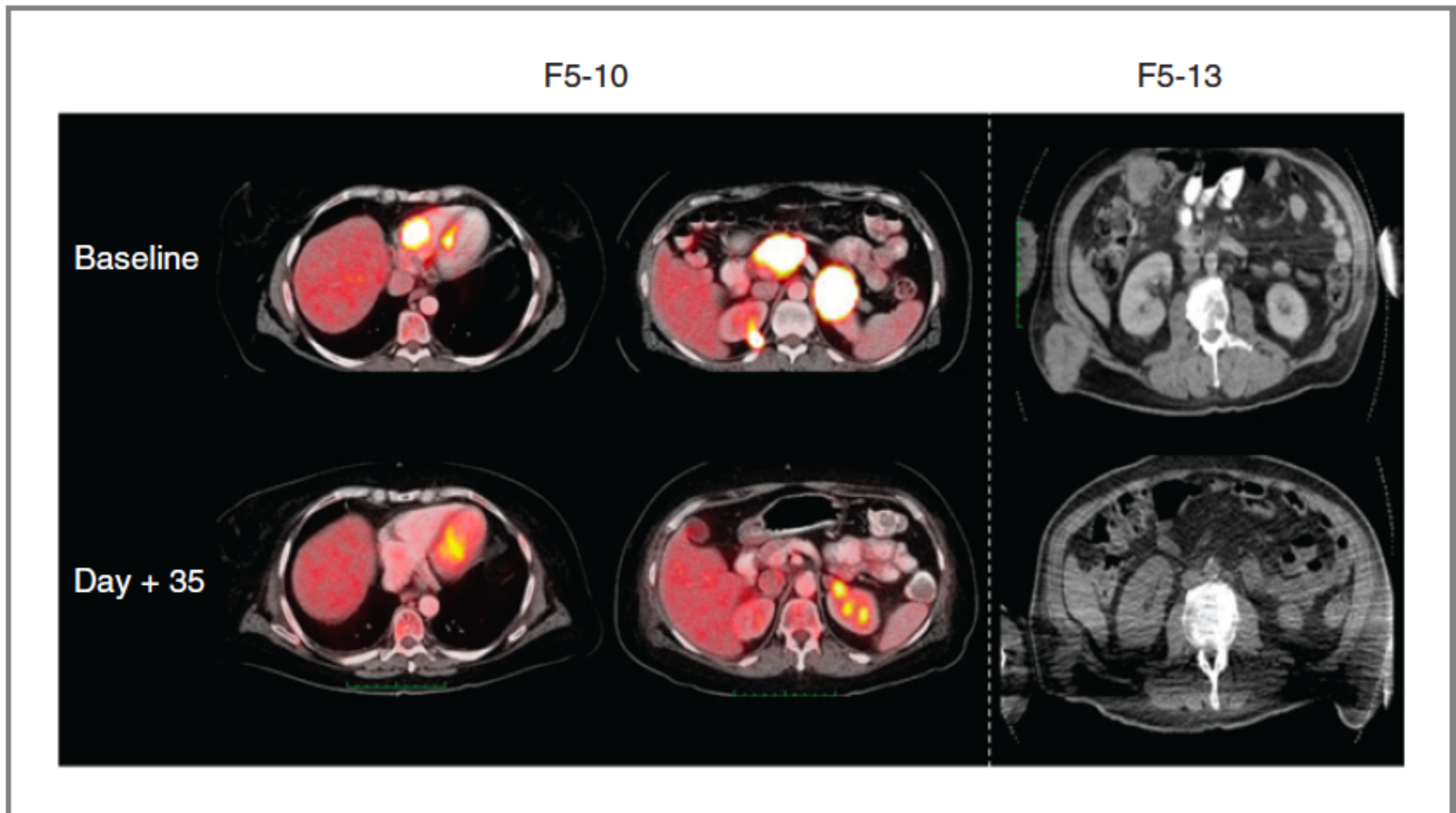
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(Johnson et al., Blood 2009)
- 2014: MART-1 TCR gene therapy + DC vaccination
 - Response in 11/14 (not according RECIST)
 - SD at 90 days in 50%
(Chodon et al. Clin Cancer Res 2014)

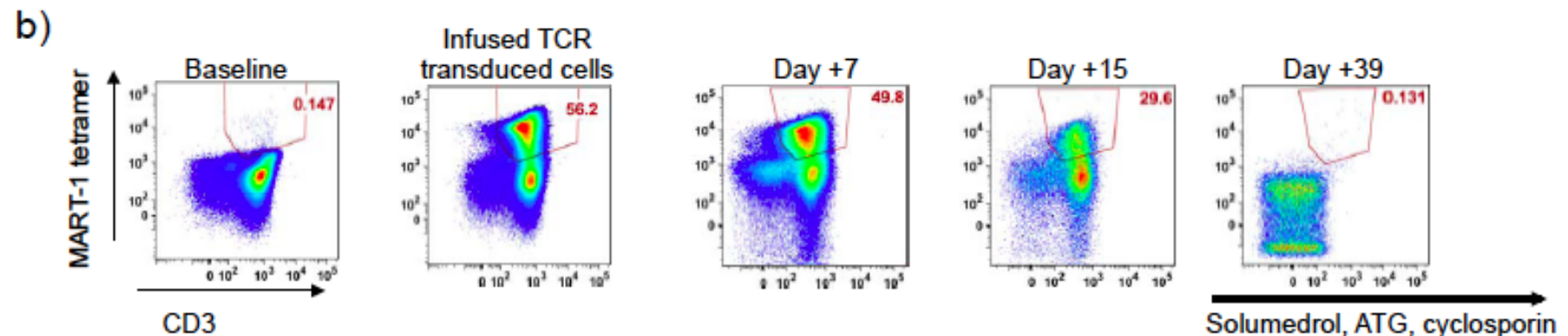
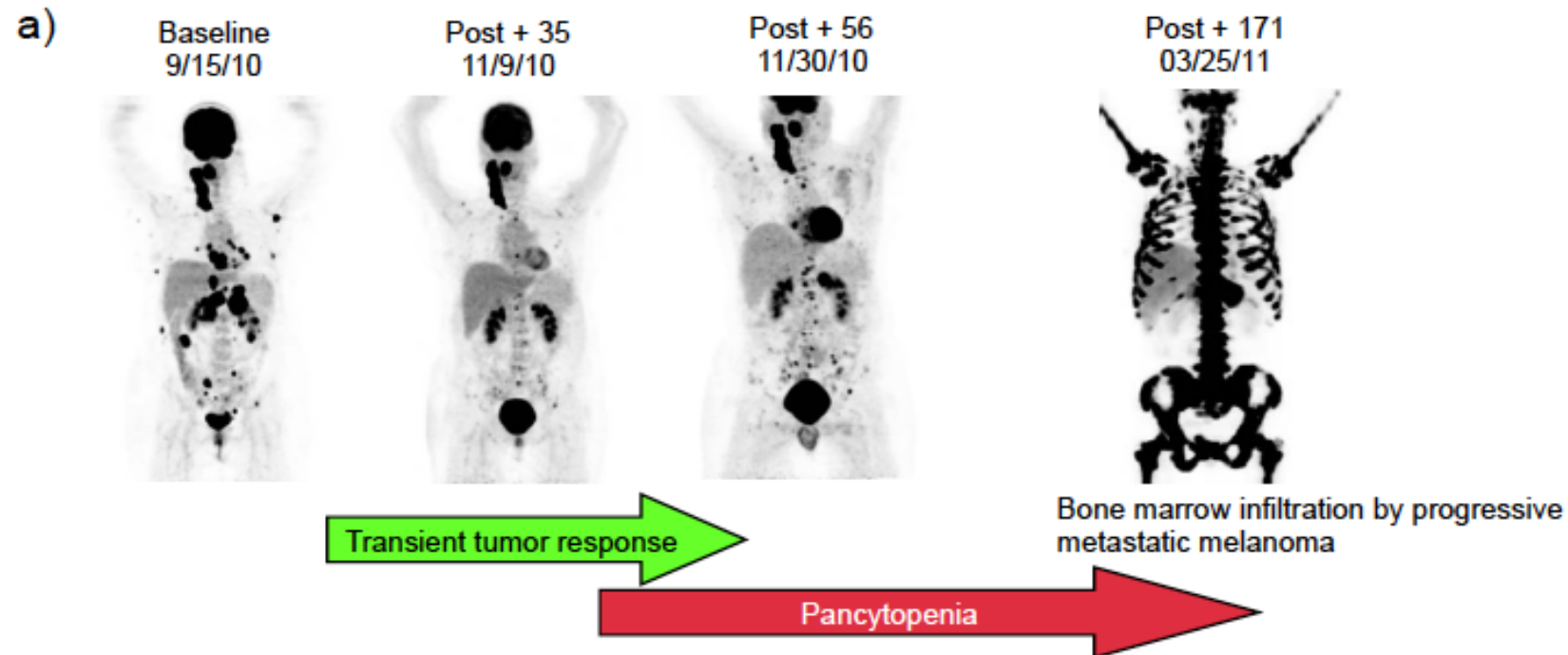
Persistence of gene modified (non-cryopreserved) cells after infusion



Clinical responses upon adoptive T-cell transfer



Pre- and post-treatment PET scans showing evidence of major tumor response and MART-1-specific TCR transgenic cell levels in patient F5-10



TCR gene therapy for melanoma

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 - RR 13% (n=15)
 - (Morgan et al., Science 2006)
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 - Response in 11/14 (not according RECIST)
 - SD at 90 days in 50%
 - (Chodon et al. Clin Cancer Res 2014)
- 2012: MART-1 TCR gene therapy (Haanen et al. unpublished)

Clinical ACT program NKI-AVL

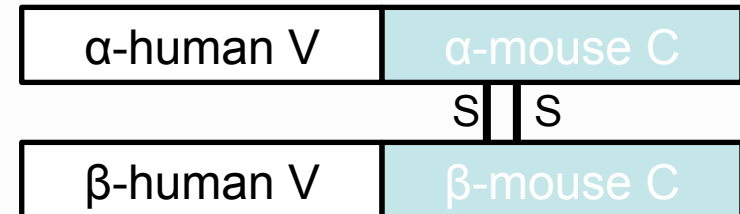
- TIL treatment melanoma
 - Current status: 10 patients treated in phase I/II feasibility trial
 - Future: Randomized multicenter phase III trial
 - TIL treatment vs ipilimumab as first or second line treatment
 - In collaboration with:
 - Herlev Hospital, Copenhagen (Inge Marie Svane & Marco Donia)
 - University of Manchester (Robert Hawkins & Ryan Guest)
- MART-1 TCR transduced T cells
 - Phase I/II trial in collaboration with Univ of Lausanne

TCR choice and expansion protocol

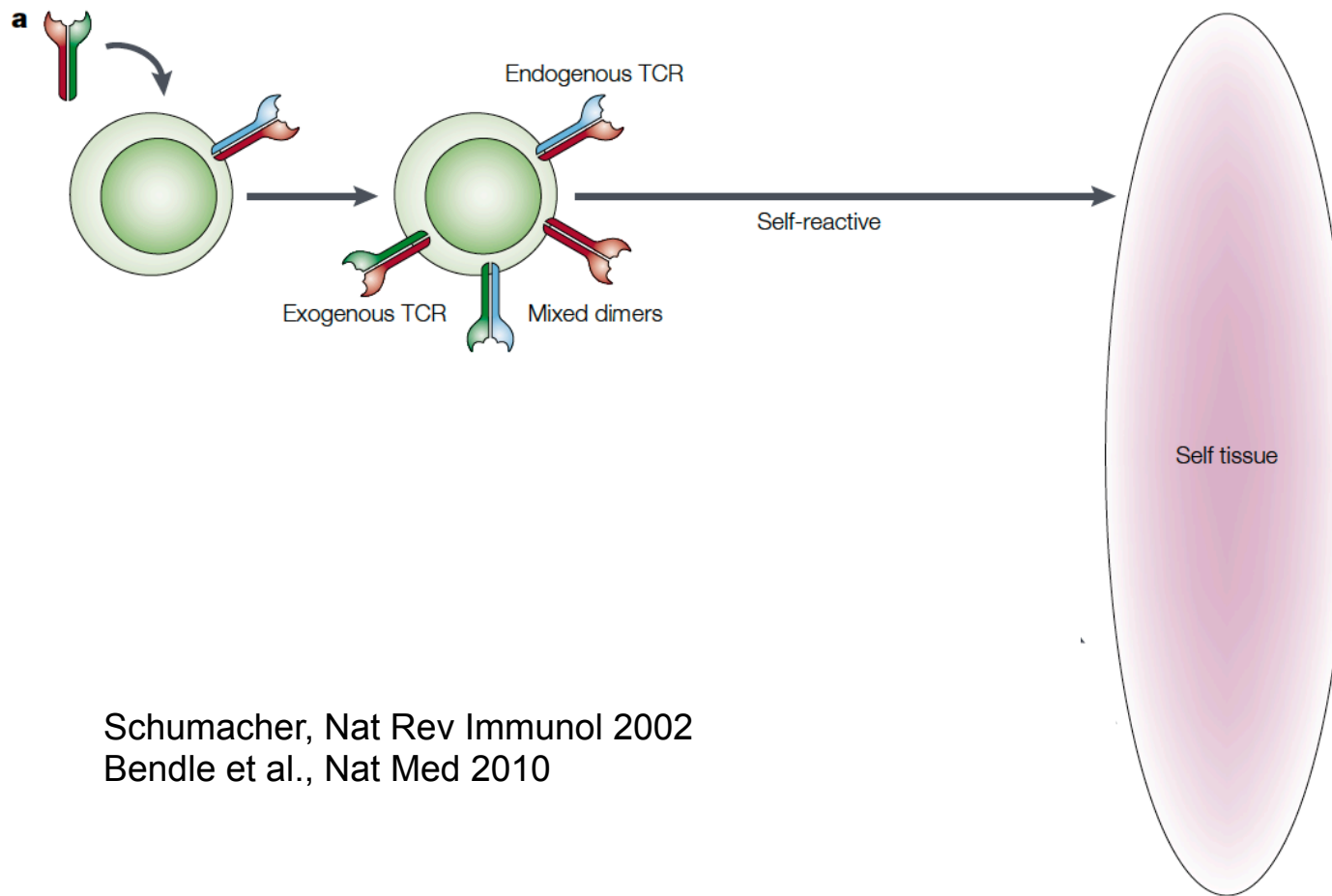
Design 1D3_{opt}HMCys TCR construct:

- 1D3 TCR recognizes MART-I 26-35 epitope (not affinity-matured)
- MP71 retroviral vector

(Jorritsma & Gomez-Eerland et al. Blood 110(10):3564-72, 2007)



Potential toxicities of TCR gene therapy (I)



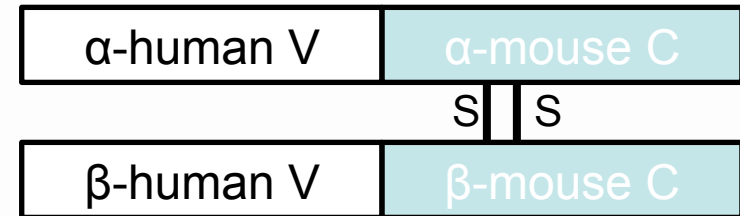
Schumacher, Nat Rev Immunol 2002
Bendle et al., Nat Med 2010

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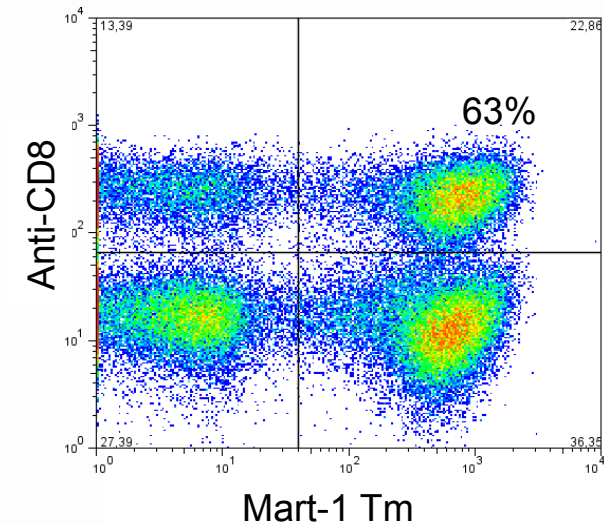
(Jorritsma & Gomez-Eerland et al. Blood 110(10):3564-72, 2007)



Expansion IL-7/IL-15 + aCD3/aCD28 beads

- “Less differentiated” phenotype compared with IL-2 + aCD3 mAb
- Better engraftment in humanized mouse model

(Kaneko et al. Blood 113(5): 1006-15, 2009)



Trial design

Patient group: Stage IIIC/IV melanoma

Clinical protocol: Simon 2-stage design phase I/II study

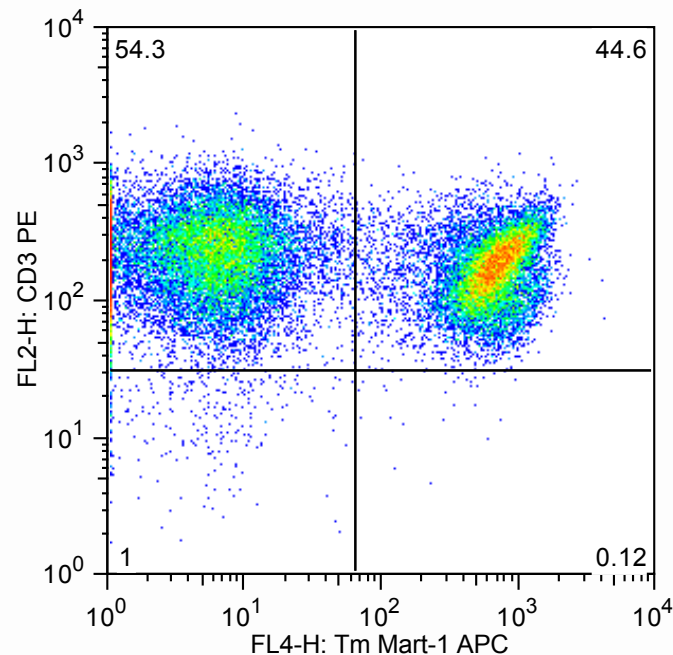
- Non-myeloablative chemotherapy cyclophosphamide/fludarabine
- T cell infusion (5×10^9 cells)
- Low-dose interleukin-2 (2×10^6 IU/once daily up to two weeks)

Clinical results so far...

- 3 patients have been treated
 - Patient 1 died at day 9 following T cell infusion
- Protocol was put on hold and was amended (100x fewer cells to be infused)
 - Two patients were treated
 - Patient 2: mixed response (PD)
 - Patient 3: 1st evaluation CT in 1 week

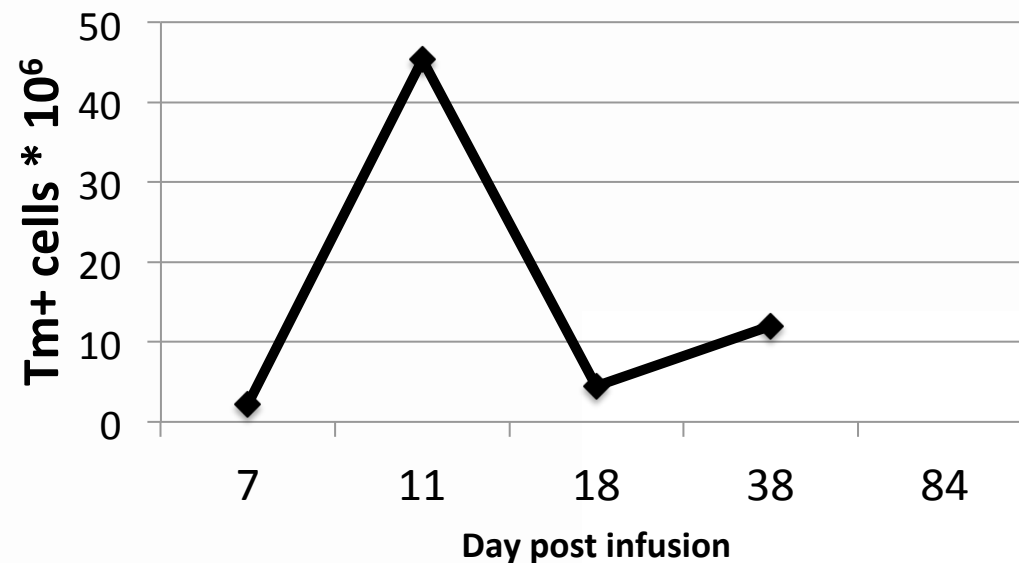
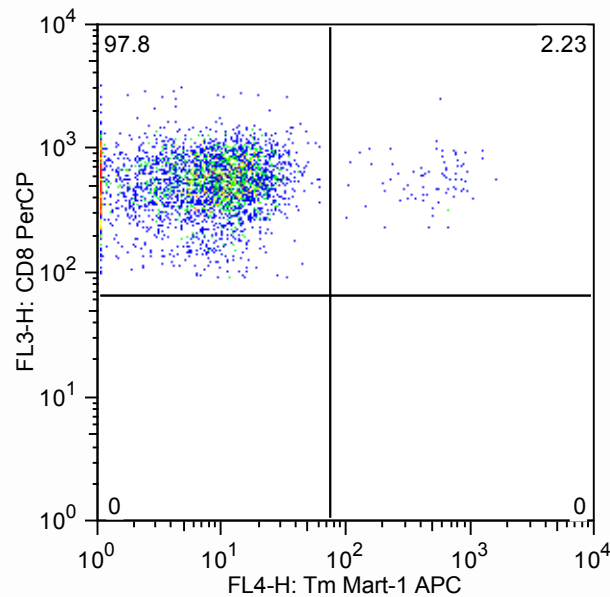
Patient 2

- 48 year old man
- Liver, cutaneous, kidney , lung and bone metastasis
- Heavily pretreated
- 5×10^7 transduced cells



Patient 2

- No severe toxicity
- Stable levels of IL-6, LDH, CRP, pro-calcitonin (inflammation markers)
- Blood samples showed the presence of modified T cells



Calculation based on the amount of lymphocytes measured by the AKL/ml and for ~ 5 L blood per person

Patient 2

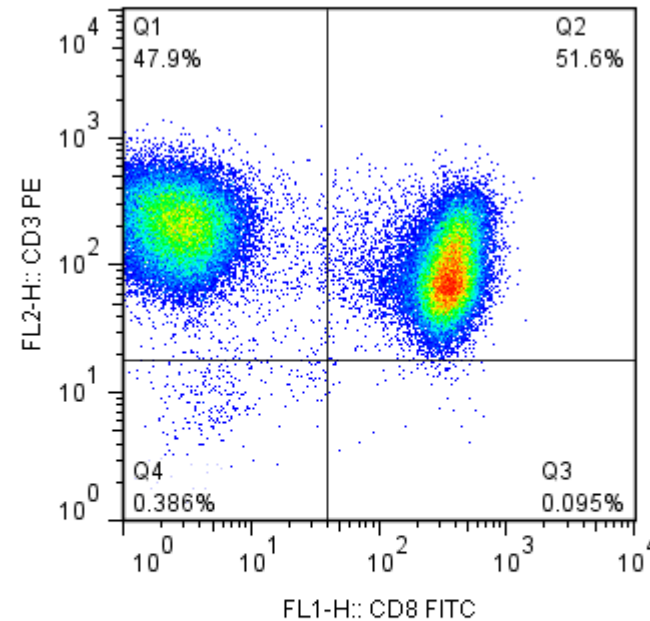
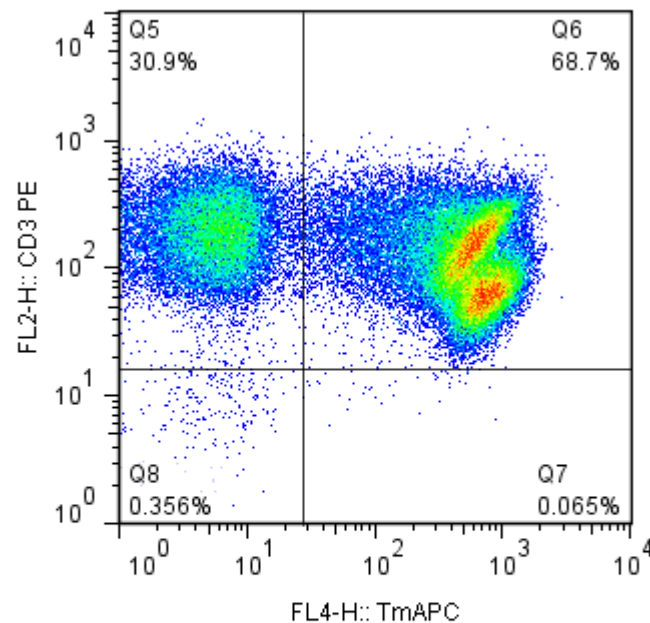
- Regression of some subcutaneous metastases
- On CT-scan: progression of LN and bone metastases at 4 weeks after infusion
- 2 months after infusion: surgery because of pathological fracture: tumor material was negative for MART-1

Patient 3

- 75 year old woman
- Metastasis in lymph node, lung, spleen and liver (positive for MART-1)
- History:
 - 2009: Diagnosed with metastatic melanoma
 - 2010: Dacarbazine
 - 2011: MART-1 DNA vaccine
 - 2011 stable disease
 - 2012 progressive disease – MEK162
 - 2012 progressive disease – inclusion M11TCR trial
 - TCR trial on hold because of patient 1
 - 2013 ipilimumab
 - 2014 progressive disease – re-inclusion M11TCR trial

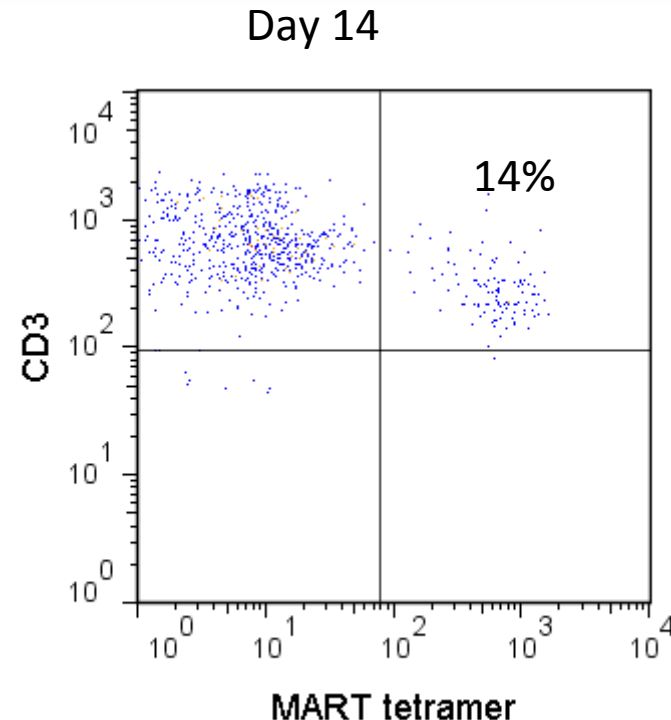
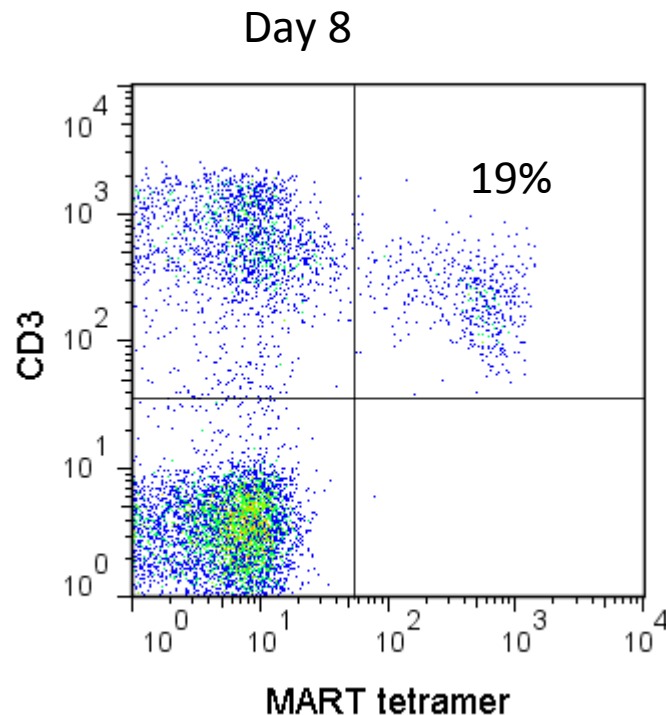
Infusion product

- 67% transduction efficiency
- 5×10^7 transduced T cells
- 52% CD8



Expansion T cells in circulation

- Absolute number in blood (5L) is estimated to be 8×10^7 on day 8



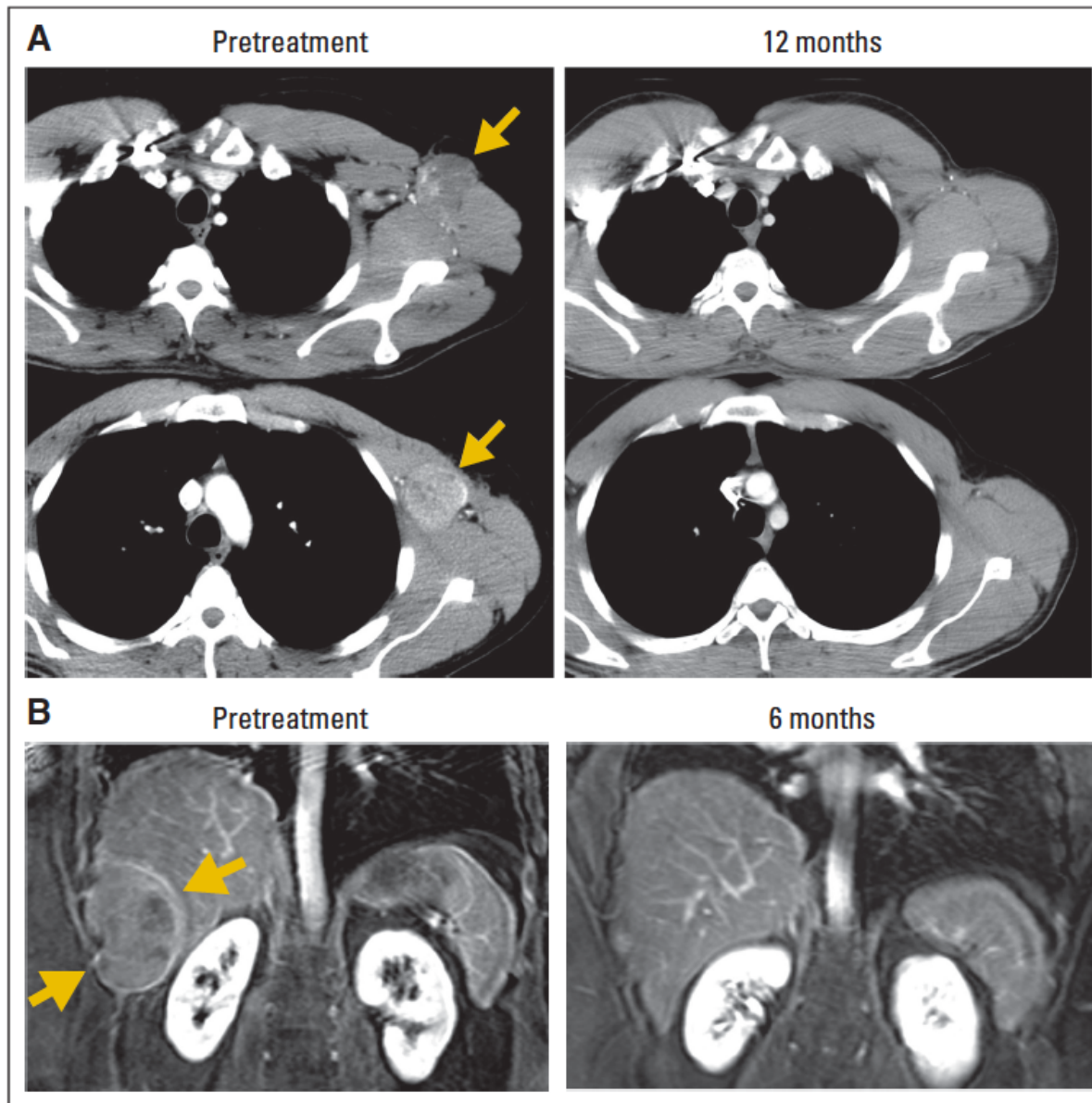
Clinical experience with TCR gene therapy

- 2006-2014: MART-1 and gp100 TCR gene therapy
- 2011: NY-eso-1 TCR gene therapy in melanoma and synovial sarcoma
 - RR 45% (n=11) and 67% (n=6)
(Robbins et al., J Clin Oncol 2011)

Patient characteristics and outcome

Patient No.	Age (years)	Sex	Sites of Disease	Prior Treatment	No. of Cells (×10 ⁹)	No. of IL-2 Doses	% of CD3		NY-ESO-1 Tetramer Positive		Vβ13.1 Positive (% of CD3)	Tumor Cell Targets (pg/mL IFN-γ)*		Response†
							CD8	CD4	% of CD8	% of CD4		NY-ESO-1 Positive	NY-ESO-1 Negative	
Melanoma														
1	52	M	ln	R, S, I	130	6	97	2	86	64	94	515	< 30	PR (8)
2	60	F	sc, lu	S, I	71	6	82	17	76	53	90	3,890	< 30	PD
3	30	F	bo, ln, panc, sb	R, S, I	47	1	98	1	80	65	91	11,978	130	PD
4	56	M	lu, ki	R, S, I	50	7	91	9	80	74	94	11,230	< 30	CR (22+)
5	32	M	ln	S, C, I	64	4	98	2	85	76	94	26,019	288	CR (20+)
6	38	M	ln	S, I	51	7	93	7	87	79	94	28,907	536	PR (3)
7	47	M	ln, lu	R, S, I	23	7	96	4	70	58	90	9,577	178	PD
8	39	F	ln, br, lu	R, S, C, I	38	8	68	32	78	70	94	ND	ND	PD
9	51	F	lu, ln, li	S, C, I	31	10	94	6	83	69	96	11,952	35	PD
10	61	M	ln, li, spl, lu, bo	R, S, C, I	16	8	84	16	79	56	92	16,063	49	PD
11	46	M	lu, li	R, S, I	37	6	93	7	63	58	85	5,795	< 30	PR (9+)
Synovial cell sarcoma														
12‡	20	M	lu, bo	R, S, C, I	83	5	82	8	77	64	91	10,065	117	PR (10)
13‡	37	F	lu	R, S, C	50	8	90	5	78	78	93	11,656	94	PR (18)
14‡	47	F	lu, ln	R, S, C	56	8	89	11	81	76	91	10,836	50	PR (5)
15‡	19	M	lu	R, S, C, I	16	5	46	40	67	63	89	5,371	< 30	PD
16	30	M	pl, hi	S, C	59	5	92	8	74	57	88	6,512	199	PR (8)
17	40	M	pl, hi	R, S, C	52	5	81	18	78	69	92	8,098	< 30	PD

Clinical responses



Clinical experience with TCR gene therapy

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 - RR 45% (n=11)
(Robbins et al., J Clin Oncol 2011)
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 - Response in 2/3 patients (1 according RECIST)
(Parkhurst et al., Mol Therapy 2009)

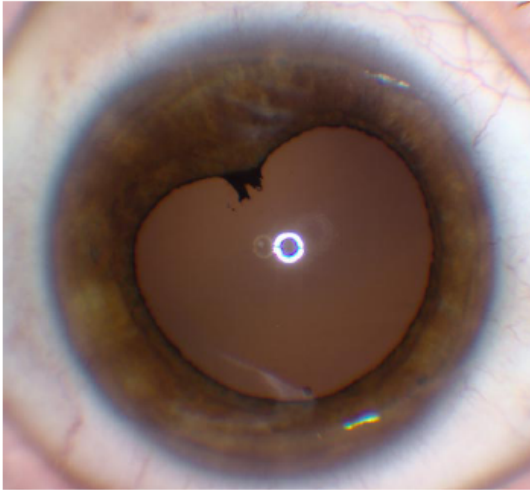
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 - Response in 2/3 patients (1 according RECIST)
(Parkhurst et al., Mol Therapy 2009)
- 2012 MAGE-A3 TCR gene therapy
 - Two independent trials were aborted due to unexpected toxicity
(Morgan et al., J Immunother 2013; Linette et al., Blood 2013)

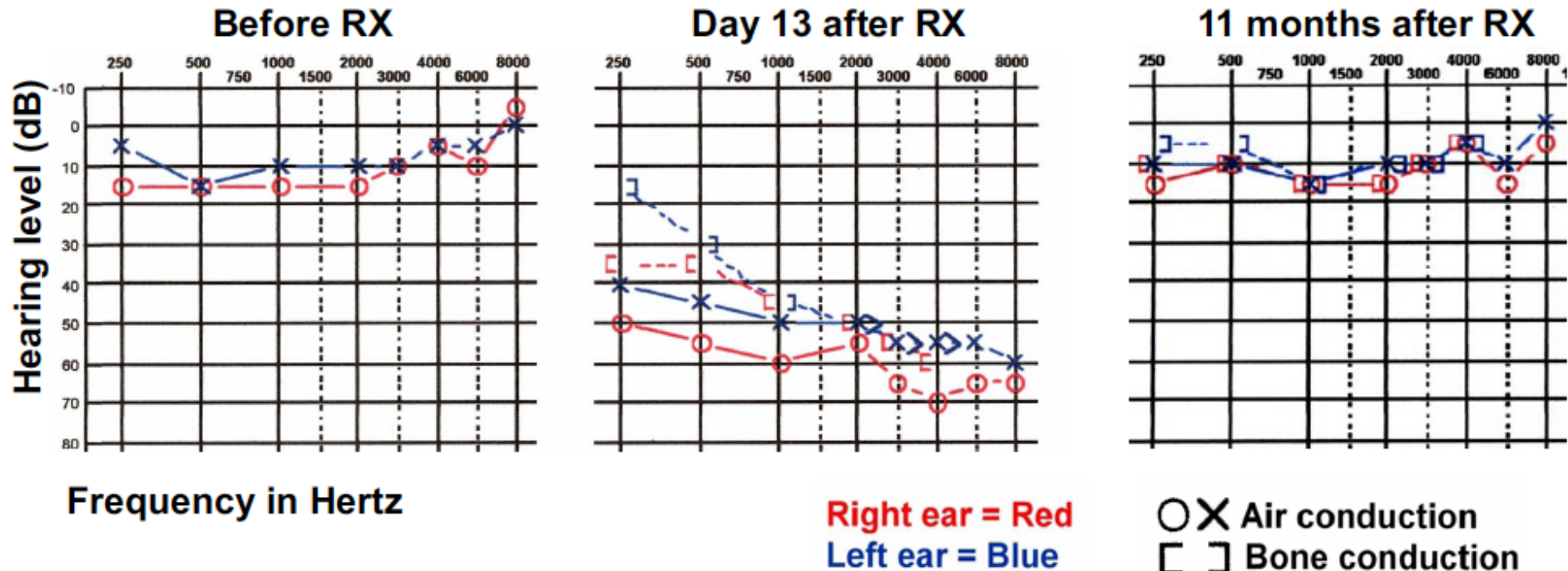
Toxicity observed in TCR gene therapy trials

- DMF4: no toxicity reported
- DMF5 + gp100 TCR:
 - Skin rash (100%)
 - Uveitis (50% of DMF5 treated pts)
 - Hearing loss (Vogt-Koyanagi-Harada) 25%
- DMF5 + DC vaccination
 - Skin rash and cytokine release syndrome
- 1D3 TCR
 - Cytokine release syndrome (multi-organ failure)
 - Skin rash

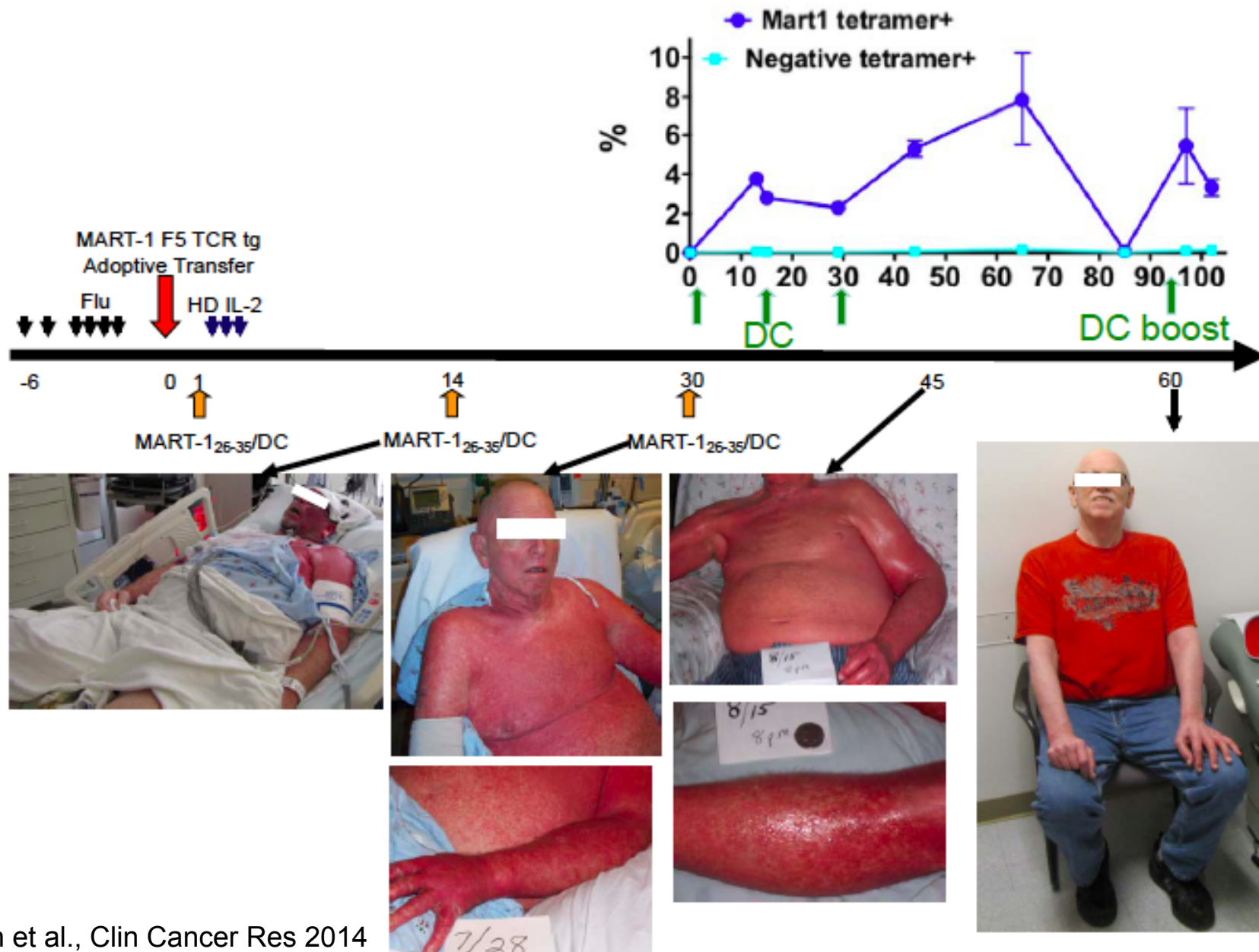
On target toxicity in eye and inner ear



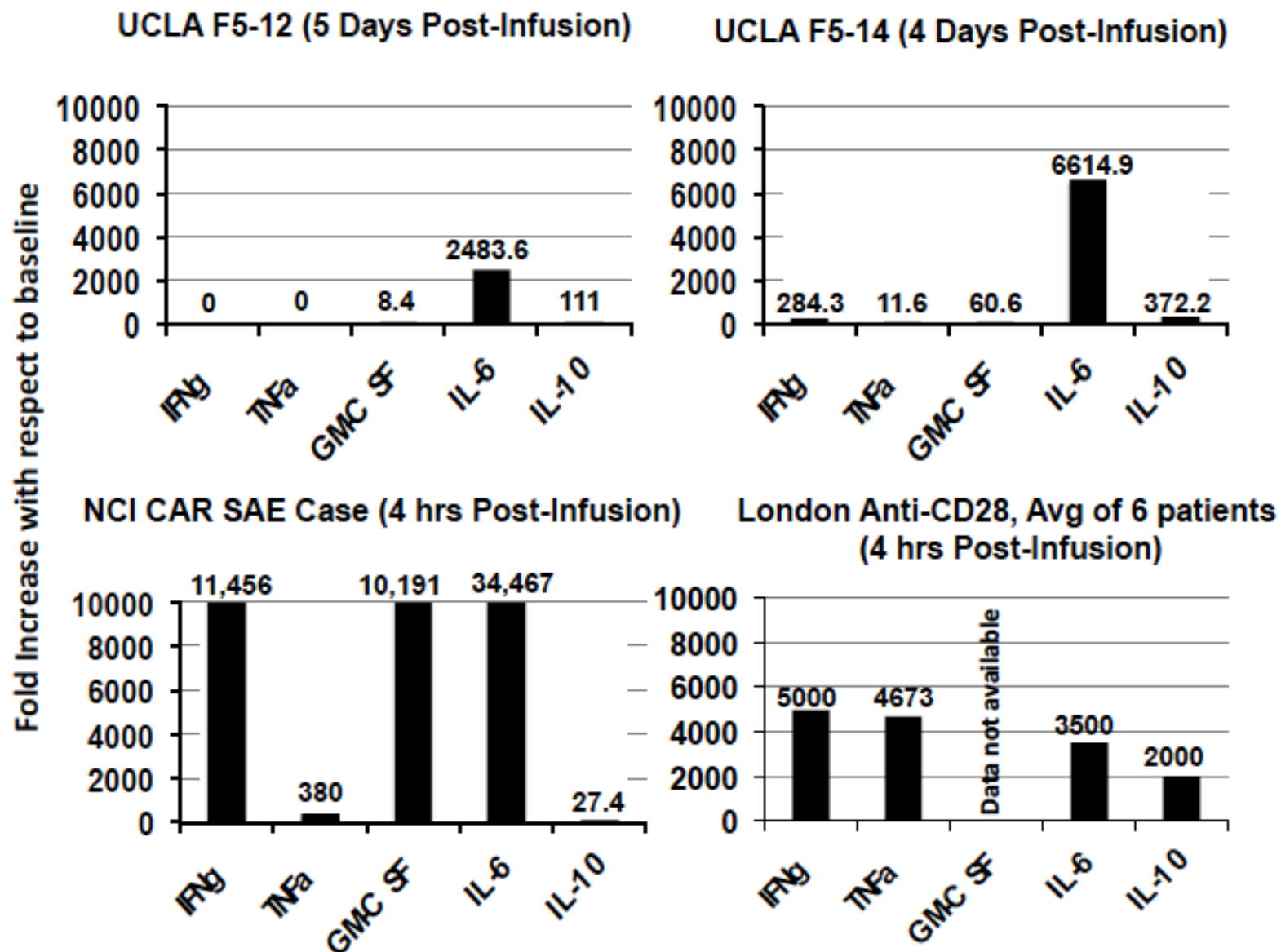
Johnson et al., Blood 2009



Recall whole body rash and re-expansion of the TCR transgenic cells in peripheral blood of patient F5-13 with subsequent MART-1/DC vaccination



Cytokine production by multiplex assay in plasma from patients F5-12 and F5-14 to study the potential development of a cytokine storm

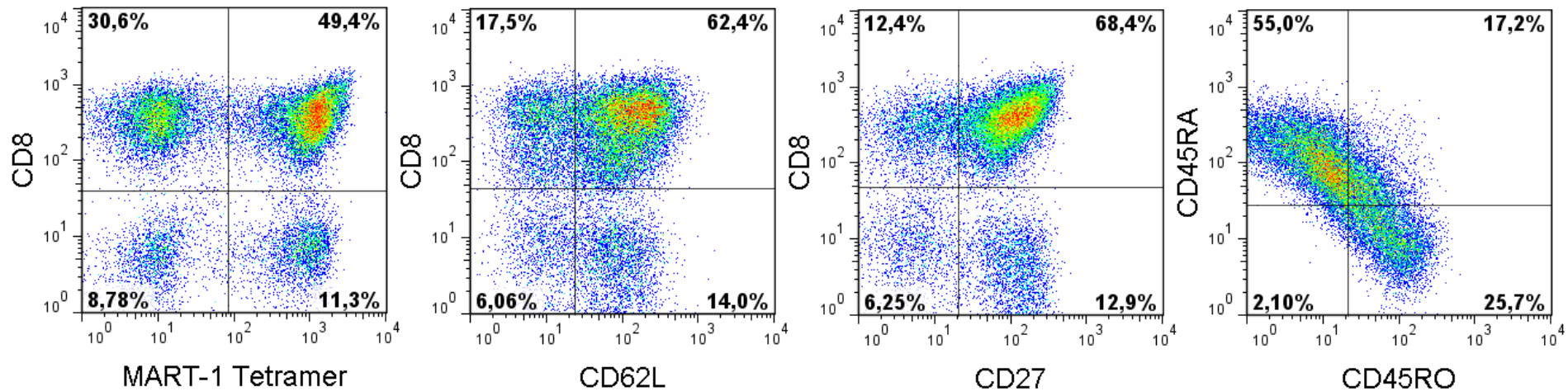


Patient 1

- 43 year old female
- Bulky disease with two large abdominal metastases (16 and 18 cm)
- Multiple pulmonary, subcutaneous and lymph node metastases of 1-3 cm
- One small brain metastasis of 8mm
- Large volume (~10L) ascites

Infusion product: characteristics

- Transduction efficiency 61%
- 4.56×10^9 transduced cells
- CD8⁺/CD4⁺ ratio 80/20
- Sterile
- Functional *in vitro*



Clinical course

- T cell infusion at day 0
- At day 1, patient developed high fever ($>40^{\circ}\text{C}$)
- Blood positive for ESBL at day 1
- Patient treated with imipenem and vancomycin
- Stabilized during following days, no signs of sepsis
- Vancomycin discontinued at day 4
- Patient showed fluid retention
- No sign of on-target toxicity against MART-1

Clinical course

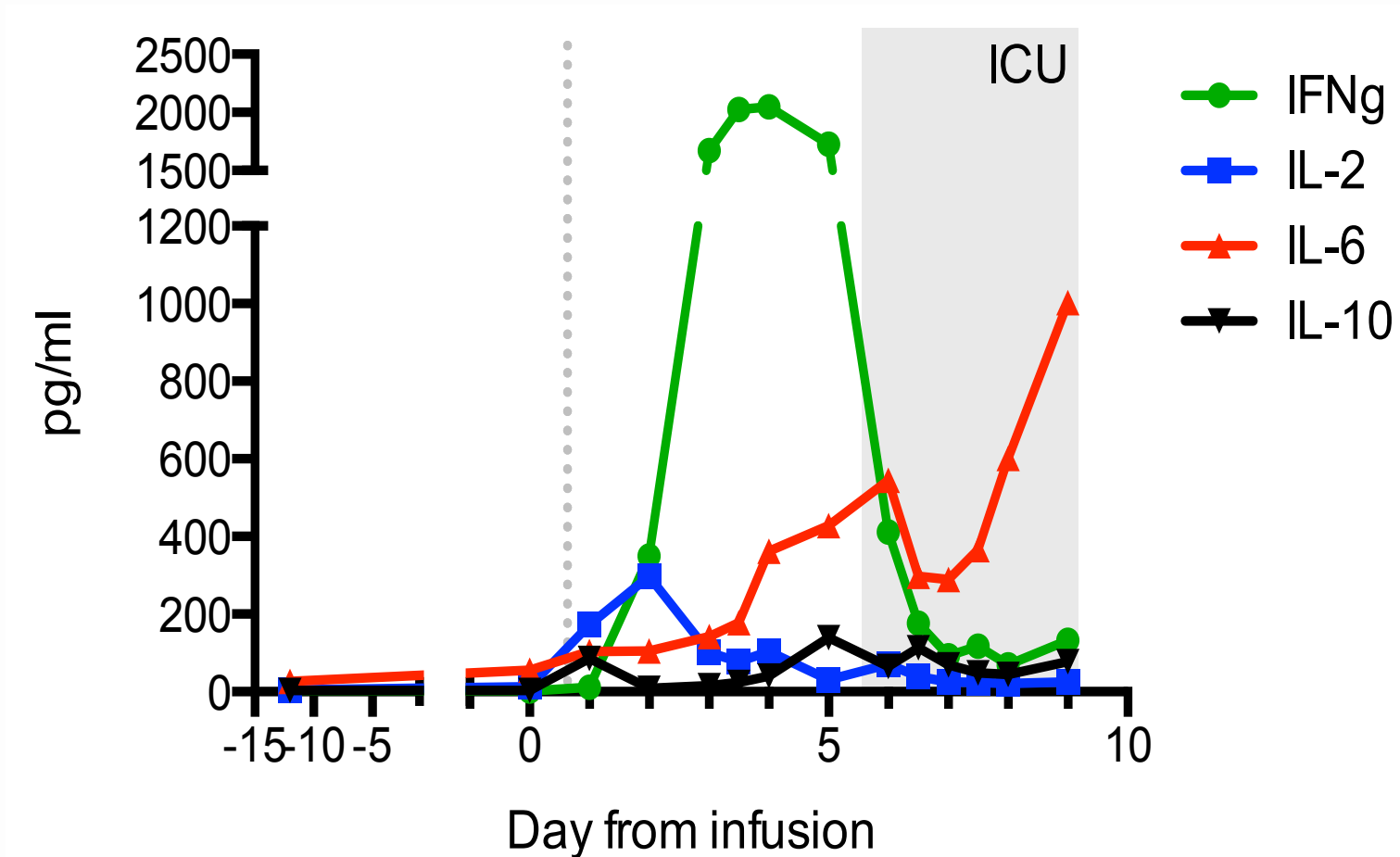
- Morning of day 6, not responding to verbal stimuli
- Fever of 39.5°C, high pulse (110/min)
- Antibiotic regimen changed to meropenem, vancomycin and amoxicillin
- Transport to ICU. Generalized tonal clonal convulsion.
- Cardiac arrest
- Intubated and resuscitated for 10 minutes
- Kept in coma
- CT scan showed brain edema and bleeding of lesion
- All cultures remained sterile (sputum, BAL, ascites, liquor)
- No meningitis
- No tumor lysis syndrome
- After 48 hours, sedation stopped, no neurological improvement → patient died on day 9 from multiple organ failure

Possible explanations

1. Bacterial sepsis
2. Cardiac arrest induced by heart failure/
hemorrhage
3. T cell related
 - a) Recognition other MHC-peptide complex
 - b) Cytokine release

Cytokine release in multiple organ failure patient 1 at NKI

- Consecutive rise of IFN- γ and IL-6



- Same profile BAL and ascites

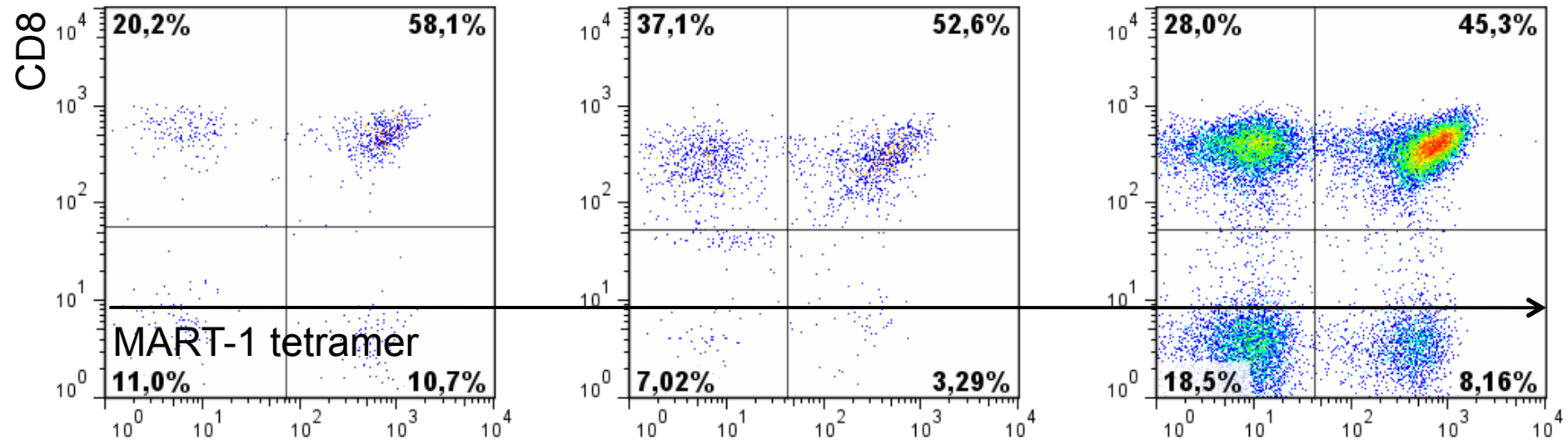
1D3 TCR modified cells present in body fluids

Day 7

PBMCs

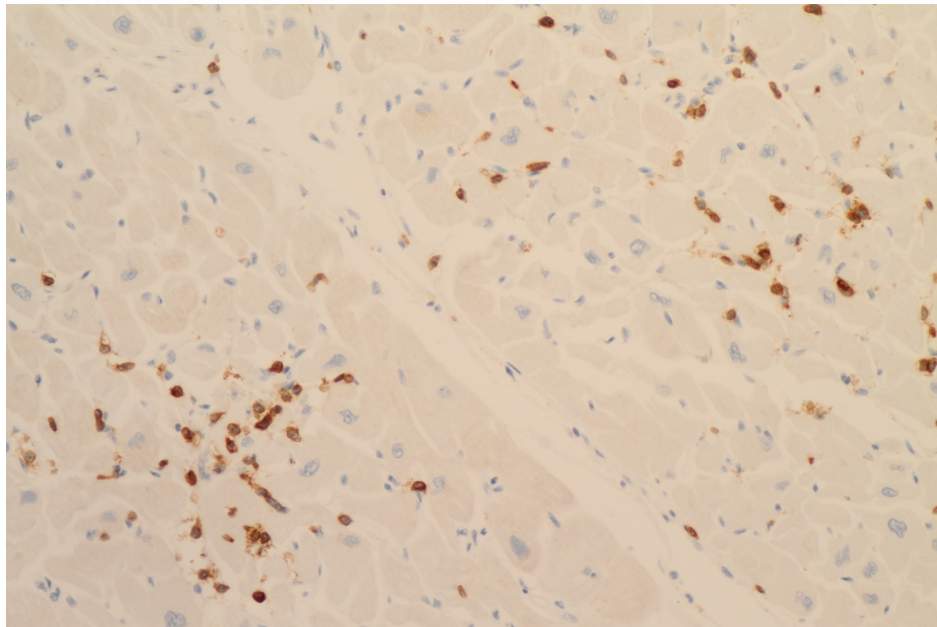
BAL

Ascites

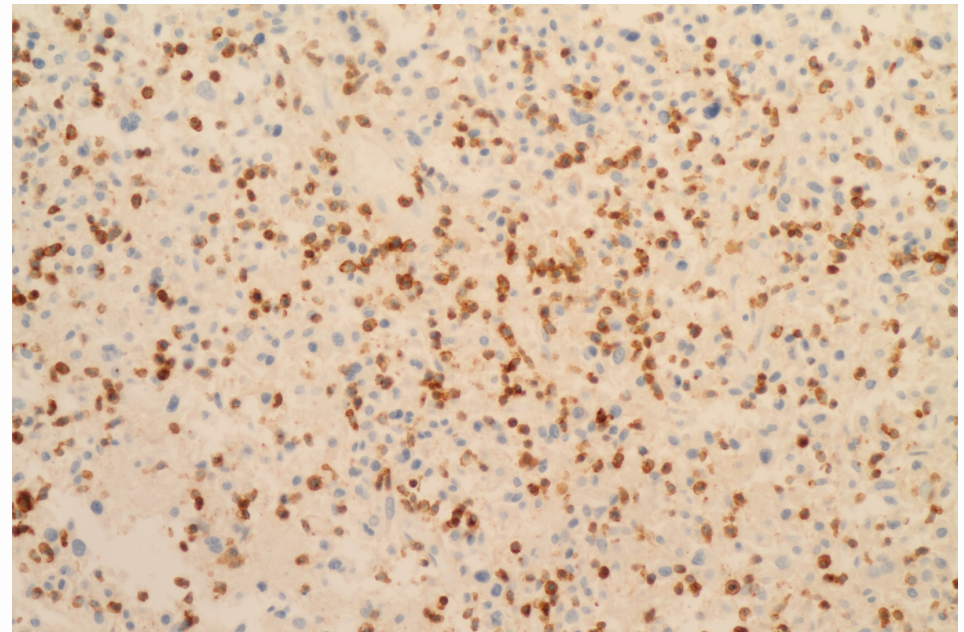


Post mortem examination

- Lymphocytic myocarditis with influx of CD3/CD8/Granzyme B/MART-1 TCR⁺ cells
- Extensive infiltration of CD3/CD8/Granzyme B/MART-1 TCR⁺ cells in peritoneal metastases
- Minor T cell infiltration in other organs



Myocardium CD3

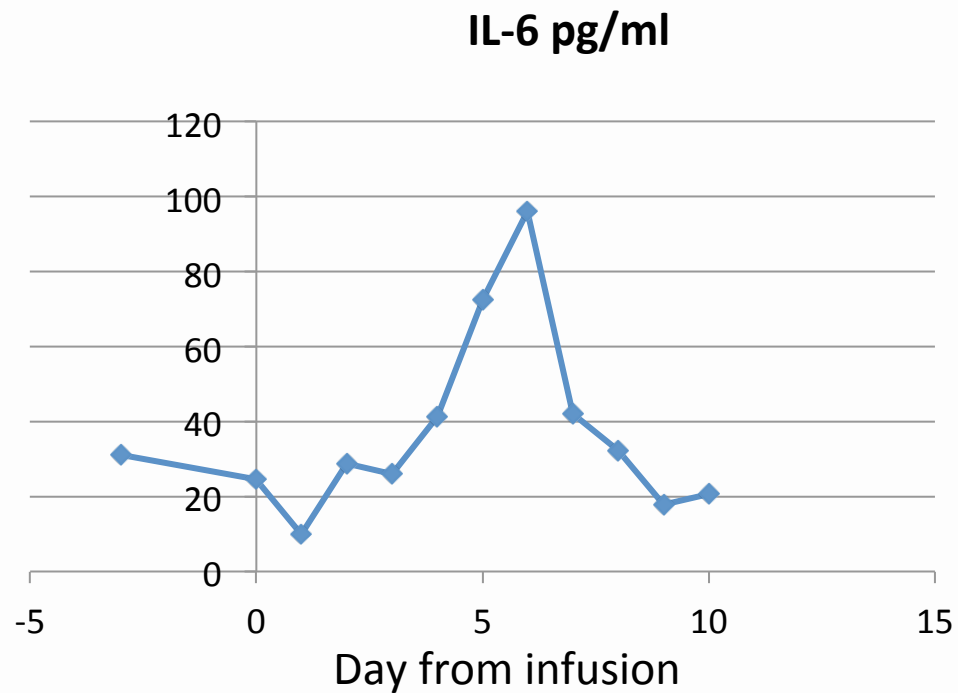


Metastases CD3

Patient 2: On-target activity

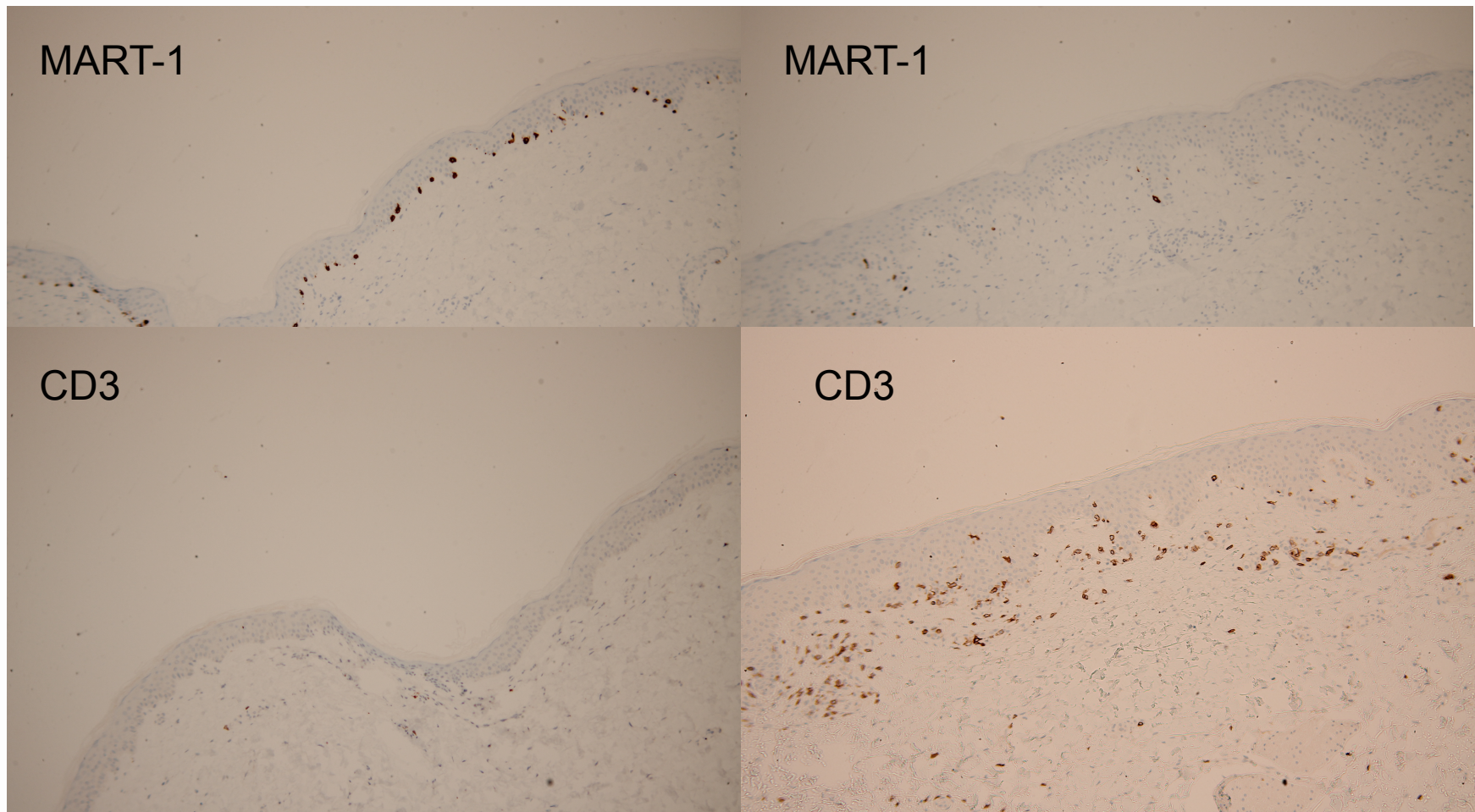
- Inflamed moles and a macular rash
 - Vitiligo (depigmentation) of the skin
-
- Biopsy of skin showed T cell infiltration
 - Local loss of melanocytes at inflammation site
 - Vitiligo confirmed by Woods lamp

Small peak in IL-6 at time of skin rash and fever

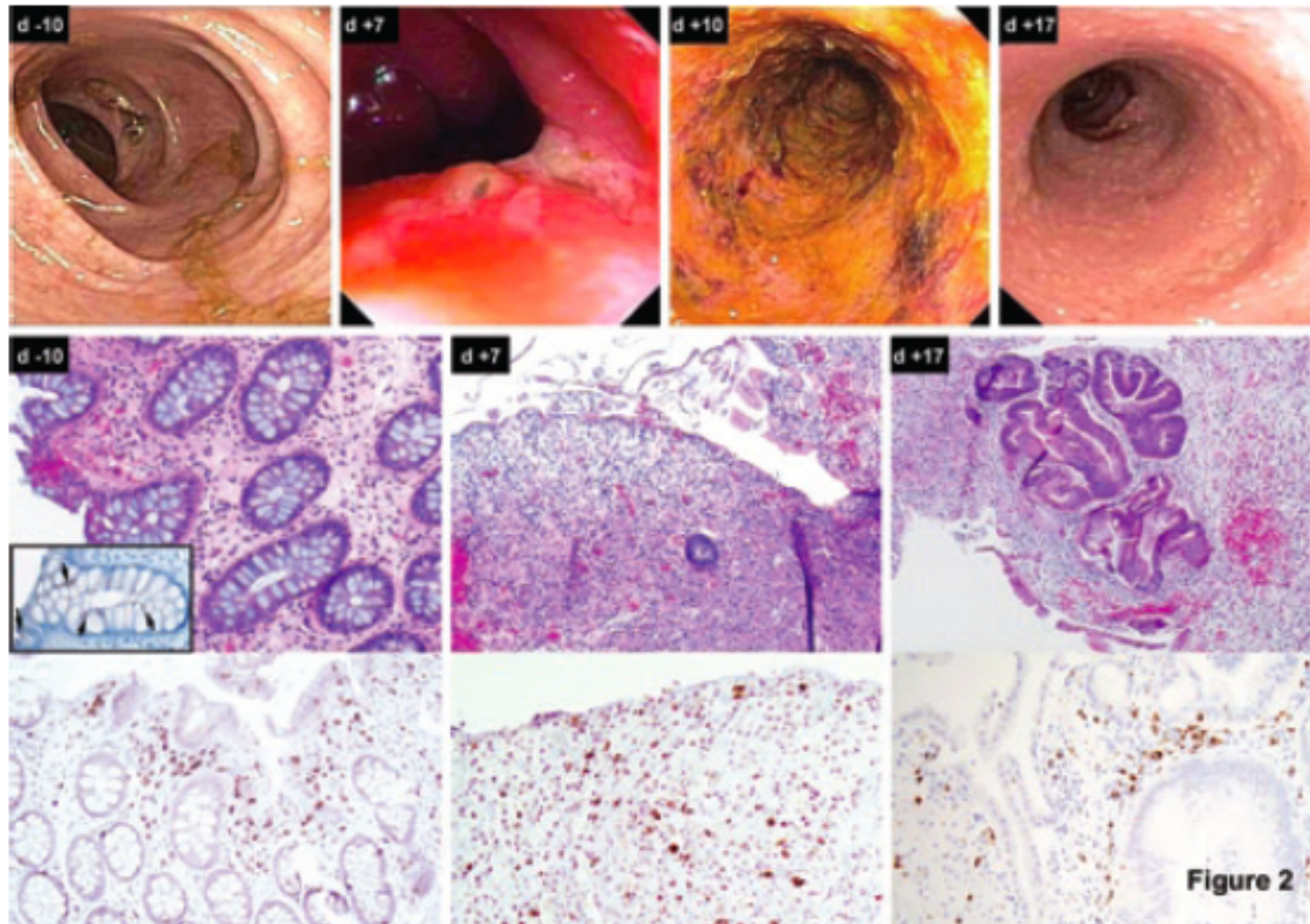


Skin reaction

- Skin rash
- Biopsy from normal skin inflamed skin



Severe colitis in CEA TCR gene therapy



Parkhurst et al., Molecular Therapy 2009

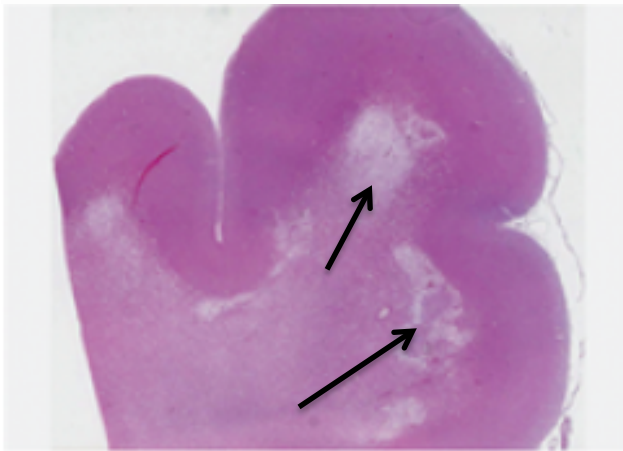
MAGE-A3 TCR gene therapy trials

- TCR recognizing HLA-A1 restricted MAGE-A3 peptide
 - Melanoma and multiple myeloma patient
 - Affinity of original TCR was improved (CDR3 region)
 - Both patients developed hypotension and severe cardiac problems leading to death within 5 days after adoptive therapy

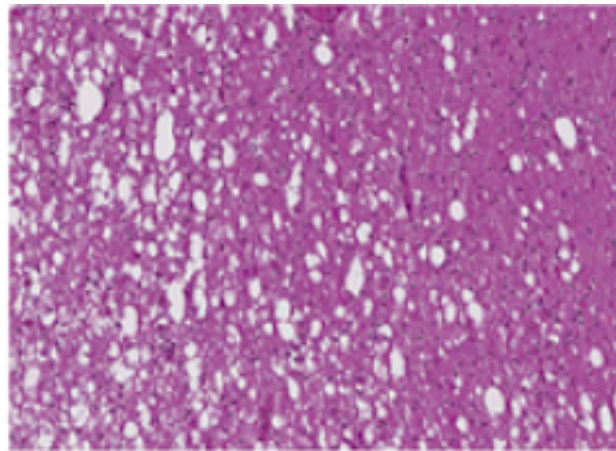
MAGE-A3 TCR gene therapy trials

- TCR recognizing HLA-A2 restricted MAGE-A3 peptide KVAELVHFL (shared with MAGE-A9)
 - 9 cancer patients expressing MAGE-A3
 - Affinity modified TCR from murine origin
 - 3 out of 9 patients developed severe neurological symptoms
 - Epileptic seizures, mental disturbances
 - Infarcted areas on brain MRI
 - 2 patients died, 1 fully recovered

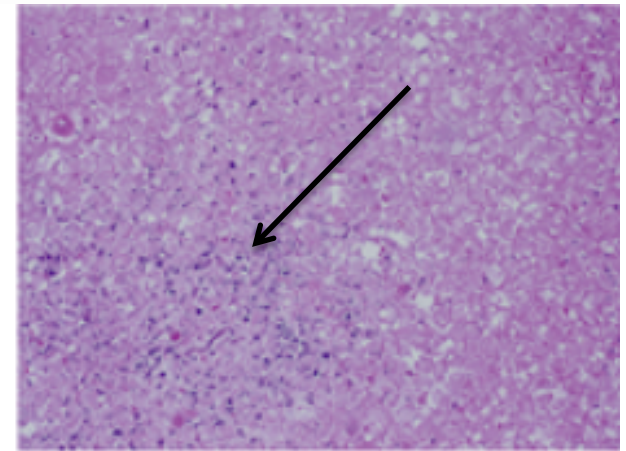
Brain damage on autopsy



White matter changes

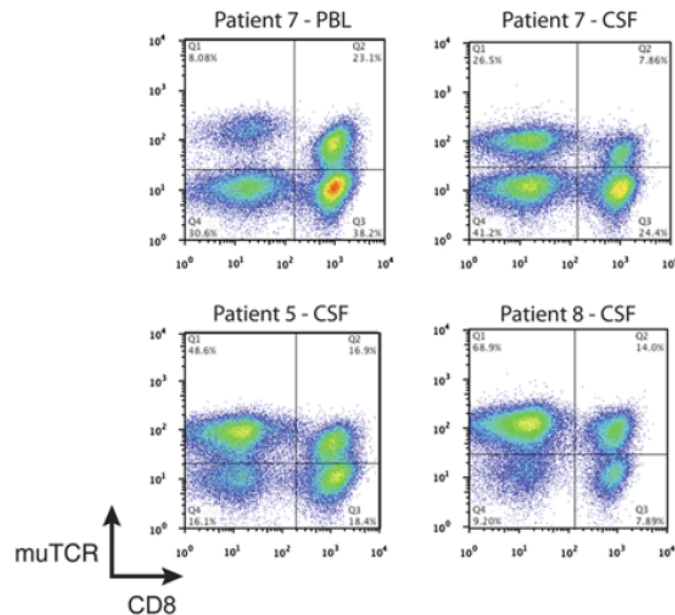


Vacuolization



Areas of infarction

Cross-reactivity of HLA-A2/MAGE-A3 peptide specific TCR with low level MAGE-A12 expression in brain



- MAGE-A12 is expressed at low level in the brain, whereas MAGE-A3 is not
- MAGE-A3 TCR recognizes MAGE-A12 peptide
- Neurological toxicity may be caused by cross-reactivity of T cells with MAGE-A12 peptide in brain

Future of TCR gene therapy

- Target choice?
- Which viral platform?
- Affinity modification?
- Suicide switch?
- Which cell type (T cell? Which T cell? NK cell?)
- How to activate and to expand?
- How many cells should we infuse?
- Deletion of endogenous TCR?
- Deletion of inhibitory receptors?
- Improve trafficking?

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